

# Chapter 3

## Intertidal Death Assemblages as Proxies of Marine Biodiversity. An Example from Northern Patagonia, Argentina



Fernando M. Archuby and Roche Andrea

1 **Abstract** Marine conservation biologists have identified mollusks as one of the  
 2 appropriate surrogate taxa for characterizing marine benthic diversity. In turn, AQ1  
 3 live/dead comparison studies have overwhelmingly demonstrated that mollusk  
 4 remains are faithful proxies of the mollusk composition of the living communities  
 5 from which they come, with positive consequences for the paleoecological evaluation  
 6 of fossil assemblages. In this contribution, we evaluate the way in which mollusk  
 7 biodiversity is distributed along the lower intertidal to supratidal (high water mark)  
 8 dead shell assemblages accumulated on a northern Patagonian rocky shore, in order  
 9 to explore the usefulness of these assemblages as paleontological proxies and potential  
 10 surrogates of regional biodiversity. A diversity gradient from the lower intertidal  
 11 to the supratidal was identified which is probably associated with vertical transport,  
 12 although the influence of gradients of the living community should be tested  
 13 to confirm this. The outstanding result of this study is the discovery of high levels  
 14 of diversity among dead shells (31 bivalves and 39 gastropod species) in a single  
 15 locality and with a moderate sampling effort. The supratidal death assemblage has  
 16 higher species richness than expected, possibly caused by stranding of the fauna  
 17 after storms. Nevertheless, this level shows the lowest level of evenness and a strong  
 18 bias when samples are not sieved through a fine mesh. The record of marine benthic  
 19 diversity in death assemblages is a promising area of research that deserves to be  
 20 explored in depth.

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F. M. Archuby (✉)  
 Instituto de Investigación en Paleobiología y Geología (IIPG), UNRN-CONICET, General Roca,  
 Río Negro, Argentina  
 e-mail: [farchuby@gmail.com](mailto:farchuby@gmail.com)

R. Andrea  
 Escuela Superior de Ciencias Marinas (ESCiMar) and Centro de Investigación Aplicada y  
 Transferencia Tecnológica en Recursos Marinos Almirante Storni (CIMAS), San Antonio Oeste,  
 Río Negro, Argentina  
 e-mail: [andrear.ibmp@gmail.com](mailto:andrear.ibmp@gmail.com)

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21 **Keywords** Intertidal · Death assemblage · Rocky-bottom · Patagonia · Depth  
22 gradient

### 23 3.1 Introduction

24 Dead shells accumulated on the sea floor contain a wealth of information which  
25 is useful either for assessing important questions of the genesis of fossil deposits  
26 (taphonomy) or for studying living communities. In the search for evidence to deter-  
27 mine how representative the fossil record is of communities that lived in the geo-  
28 logical past, the discipline of taphonomy has developed tools which provide high-  
29 quality information of living ecosystems. This is achieved by allowing their fea-  
30 tures to be explored on longer timescales, beyond those typically used by ecologists  
31 (Kidwell and Tomašových 2017; Olszewski and Kidwell 2007; Tomašových and  
32 Kidwell 2009a; Archuby et al. 2015; De Francesco et al. 2013; Yanes et al. 2008;  
33 Hassan et al. 2018). Current developments go still further: it is now possible to iden-  
34 tify the effect of human impact on ecosystems, by studying the differences between  
35 impacted living communities and time-averaged assemblages accumulated over the  
36 past decades or centuries (Erthal et al. 2011; Kidwell 2008; Yanes 2012; Dietl et al.  
37 2015). The relevance of this paleobiological information, which offers us an oth-  
38 erwise inaccessible long-term perspective of biodiversity and community change,  
39 has given rise to a new discipline: conservation paleobiology (Barnosky et al. 2017;  
40 Louys 2012; Rick and Lockwood 2013; Dietl and Flessa 2011; Dietl et al. 2015;  
41 Kidwell 2009; Kidwell and Tomašových 2013).

42 Biodiversity is of fundamental importance to ecology because it is the conse-  
43 quence of how organisms in communities respond to biotic and abiotic factors  
44 (Olszewski and Kidwell 2007). The use of biological surrogates (i.e. estimators,  
45 such as polychaetes, crustaceans, mollusks, etc.) to evaluate marine biodiversity is  
46 a useful practice in conservation biology because it helps overcome the difficulties  
47 inherent in surveying benthic communities: time and cost, the occurrence of unde-  
48 scribed species and the problems of species identification (Tyler and Kowalewski  
49 2017; Magierowski and Johnson 2006; Mellin et al. 2011; Warwick and Light 2002).  
50 Research focuses on finding appropriate surrogates for the different types of marine  
51 communities and their spatial and temporal variations. Molluscs, which are among  
52 the groups selected as appropriate surrogates of marine benthic communities, leave  
53 abundant mineralized dead remains, which have been proved to be good proxies  
54 of the communities from which they derive (Tyler and Kowalewski 2017; Kidwell  
55 2008; Smith 2005).

56 Assessing how diversity transfers from living communities (life assemblages,  
57 LAs) to death assemblages (DAs) is a crucial step towards a better interpretation of  
58 diversity in fossil assemblages; this knowledge will also help us to evaluate death  
59 assemblages as faithful proxies of living communities. The path from LAs to DAs  
60 represents the first filter that modifies diversity measurements, through differential

61 transport and destruction by waves, currents, wind and time-averaging (Archuby  
62 et al. 2015; Tomašových and Kidwell 2009a, 2010a).

63 In the absence of strong reworking of former beds, such as in the case of ravine-  
64 ment, coastal marine beds encompass a short time span and their skeletal content is  
65 considered representative of the average composition of successions of communities  
66 along hundreds or, at the most, thousands of years (within-habitat time-averaging  
67 of Kidwell and Bosence 1991; see also Fürsich and Aberhan 1990). Recently, the  
68 quantitative knowledge of the differences between death assemblages and living com-  
69 munities, and the sources of these differences has been greatly improved (Olszewski  
70 and Kidwell 2007; Tomašových and Kidwell 2009a, b, 2010a, b, 2011; and many  
71 more).

### 72 3.2 Death Assemblages, Taxonomic Diversity, 73 and Taphonomic Fidelity

74 Due to the time-averaged nature of DAs, their species composition is not particularly  
75 influenced by the short-term species composition fluctuations of living communities  
76 (Fürsich and Aberhan 1990; Tomašových and Kidwell 2010a; Archuby et al. 2015).  
77 These short-term fluctuations, such as the local extinction of the surf clam *Mesodesma*  
78 *mactroides* on the Atlantic coasts of Uruguay and Northern Argentina (Fiori and  
79 Cazzaniga 1999; Dadon 2005), might give totally different results in samples of living  
80 communities separated by only a few weeks. However, in this respect, DAs are highly  
81 informative due to their inertia in the face of such fluctuations. Compared with living  
82 assemblages, DAs which have accumulated over a few decades to several centuries  
83 are expected to have an increase in alpha diversity, a decrease in beta diversity  
84 (due to spatial mixing), reduced species dominance and increased frequency of rare  
85 species (Tomašových and Kidwell 2010a). Additionally, the ecological information  
86 of current ecosystems does not span more than a few decades into the past (Rick  
87 and Lockwood 2013). If we consider that human occupation of Patagonia dates from  
88 around 17,000–14,000 years BP (Perez et al. 2016), baseline ecological studies might  
89 fail to identify the non-impacted conditions when assessing anthropogenic impacts,  
90 since the impacts were already there.

91 In turn, death assemblages are used to characterise not only the average species  
92 compositions of source communities, but also biotic interactions such as local level  
93 predator-prey relationships (e.g., Visaggi and Kelley 2007; Yanes and Tyler 2009;  
94 Gordillo and Archuby 2012, 2014; Martinelli et al. 2013; Tyler et al. 2014; Archuby  
95 and Gordillo 2018), and to compare these along geographical gradients (e.g., Kelley  
96 and Hansen 2007; Visaggi and Kelley 2015). Quantifying predator-prey interactions  
97 in living communities implies sampling strategies that are complex and expensive,  
98 while the records from death assemblages are a significant source of information.

99 Studies on taphonomic fidelity (correlation of living and death assemblages) have  
100 been developed in marine, freshwater and terrestrial environments (e.g., Fürsich and

101 Flessa 1987; Kidwell and Bosence 1991; Yanes et al. 2008; Tietze and De Francesco  
102 2012; Terry 2010; more references in Archuby et al. 2015). Studies of marine death  
103 assemblages are abundant, although they are mostly based on soft-bottom ecosys-  
104 tems (Olszewski and Kidwell 2007; Kidwell 2013), and there are few studies of  
105 communities inhabiting rocky bottoms (Zuschin et al. 2000; Zuschin and Oliver  
106 2003; Zuschin and Stachowitsch 2007). Recently, Archuby et al. (2015) assessed  
107 the taphonomic fidelity of rocky-bottom communities along 1500 km of the Patago-  
108 nian Atlantic coast, from death assemblages collected at the high-water mark. These  
109 authors found a general agreement between life and death assemblages at the bio-  
110 geographical province level, working with non-sieved, representative samples (hand-  
111 picked along transects). Besides the regional agreement, on smaller geographical  
112 scales DAs tended to cluster together and are separated from LAs. So far, there are  
113 no detailed studies on the nature of DAs on rocky shores. A better understanding  
114 of the provenance of the diversity differences between life and death assemblages  
115 in modern environments is also crucial for correctly interpreting fossil assemblages  
116 and ecosystems (Olszewski and Kidwell 2007).

### 117 3.3 Purpose of This Study

118 In this study, we evaluate the way in which species richness and evenness of mollusk  
119 death assemblages is distributed along the depth gradient, from the lower intertidal  
120 to accumulations at the high-water mark, in Punta Mejillón, Northern Patagonia,  
121 Argentina. Our two goals are to improve the understanding of DAs as paleontolog-  
122 ical proxies and to evaluate their usefulness as surrogates of shallow benthic living  
123 communities. Punta Mejillón has little human impact due to its distance from the  
124 nearest city (the town of San Antonio Oeste, 105 km away), the difficulty getting  
125 there (sand dunes often cover the route), the need for a four-wheel drive vehicle to  
126 reach the beach, and also because it is located in a natural protected area (see The  
127 Study Area, below). We aim to determine whether the DAs coming from the same  
128 habitat but accumulated at different depths include specimens of different species  
129 in different proportions (i.e., there is diversity partitioning of DAs along their habi-  
130 tat). We also test the effect of sieving versus non sieving on species richness and  
131 evenness. Specifically, we aim to answer the following questions: (i) How does DA  
132 species composition vary along the lower intertidal to supratidal gradient? Is there  
133 diversity partitioning along the depth gradient in the rocky intertidal belt of northern  
134 Patagonia? (ii) Are death assemblages from rocky shores appropriate surrogates of  
135 benthic biodiversity in northern Patagonian shallow marine communities? Is there a  
136 horizon along the lower intertidal to supratidal belts that collects most of the infor-  
137 mation on the death assemblages? In other words, where is it best to sample? (iii)  
138 What is the effect of sieving on the biodiversity record?

## 139 3.4 Methodology

### 140 3.4.1 The Study Area

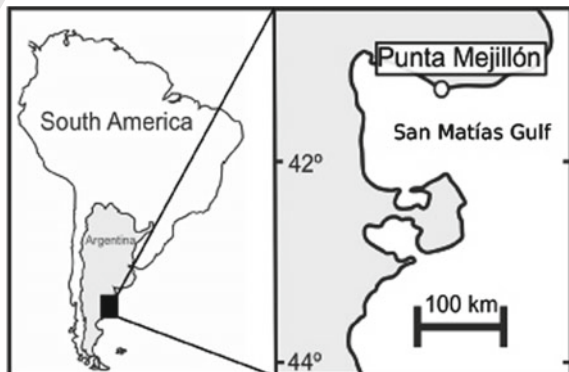
141 The study was carried out in Punta Mejillón (PM), located in the Caleta de los  
 142 Loros natural protected area in Río Negro Province, Argentine Patagonia. The place  
 143 is difficult to access, which minimizes the impact of tourism and human activities  
 144 on living communities and death assemblages (Fig. 3.1). Punta Mejillón is on the  
 145 Atlantic coast ( $41^{\circ} 00' 37''$ ) in the San Matías Gulf. The coastline runs approximately  
 146 from SE to NW, and the intertidal belt is exposed for more than 300 m during  
 147 low tides (Fig. 3.2). Biogeographically, PM is in the transition zone between the  
 148 Argentine and Magellanic Provinces and is characterized by a mixture of species  
 149 from both biogeographical entities (Balech and Ehrlich 2008). In a recent article,  
 150 Güller and Zelaya (2017) mention a surprisingly high level of mollusk diversity in  
 151 the San Matías Gulf, which they describe as a hot-spot of diversity.

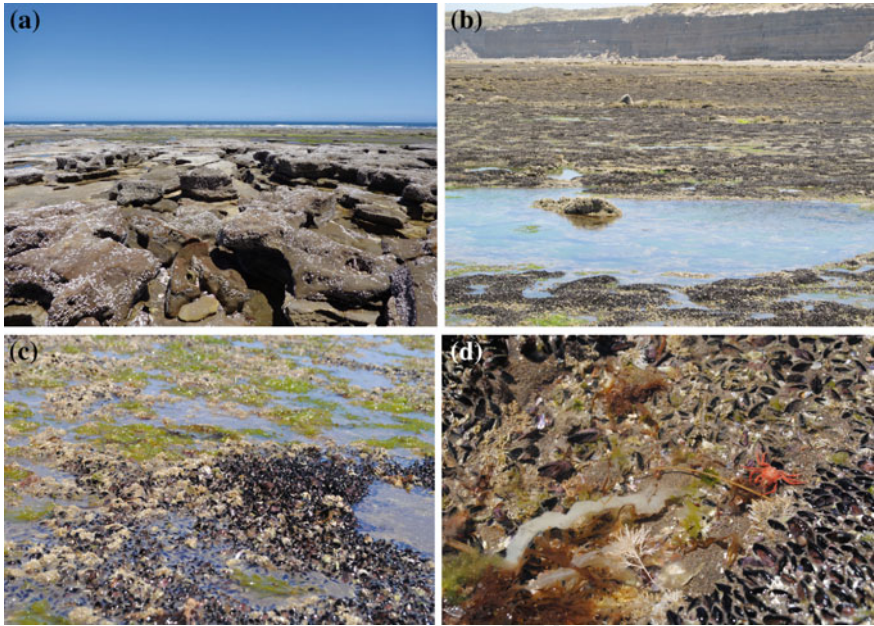
152 The northern part of San Matías Gulf, where Punta Mejillón is located, is subject to  
 153 high levels of physical disturbance, consisting of strong winds, high tidal amplitudes  
 154 (up to around 9 m) which leave large areas of the intertidal belt exposed, high-energy  
 155 flows during high tide and low temperatures (sea surface temperatures  $10.1\text{--}18.9^{\circ}\text{C}$ )  
 156 (Bertness et al. 2006; Archuby et al. 2015). Due to the high levels of desiccation  
 157 stress caused by winds, the region is considered an extremely harsh intertidal rocky  
 158 ecosystem (Bertness et al. 2006), which results in intertidal communities which are  
 159 strongly organized by physical stress.

### 160 3.4.2 Sampling

161 Sampling was carried out on 29 November 2013 during low tide, between latitudes S  
 162  $41^{\circ} 00' 32''$  and  $41^{\circ} 00' 54''$ . Samples were collected at four levels: 1. accumulation

Fig. 3.1 Map of the study area





**Fig. 3.2** Pictures of the intertidal belt in Punta Mejillón. **a** Upper intertidal. **b** View from the middle intertidal to the coast. **c** and **d** details of the middle to lower intertidal

163 of shells at the high-water mark (supratidal or “Supra”); 2. upper intertidal belt (UI);  
 164 3. middle intertidal belt (MI); and 4. lower intertidal belt (LI). At each level, several  
 165 replicates were extracted from the upper 15 cm using a shovel and were pooled  
 166 together, until completing 15 L of sediment. The replicates were extracted every  
 167 10 m along a transect parallel to the coastline. Since the substrate is mostly hard,  
 168 samples were taken from depressions filled with sediments in the area surrounding  
 169 the sampling point. In the absence of a suitable place to extract the replicate, the point  
 170 was skipped, and the sample was taken at the next point. Samples were sieved in the  
 171 field with a 10 × 10 mm aperture mesh (coarse) above and a 1 mm × 1 mm aperture  
 172 mesh (fine) below so that large shells were captured separately from small shells  
 173 (Fig. 3.3). The coarse mesh sieve retains shells that are visible and was considered  
 174 as a proxy “hand-collecting method”, that was compared with “whole” samples per  
 175 level (made by the pooling of coarse and fine samples). The 1 mm sieve was used to  
 176 explore a suitable sampling strategy for rocky-bottom dominated intertidal DAs from  
 177 the Patagonian Atlantic coast. Kidwell (2002) suggested that sampling with mesh  
 178 sizes lower than 1 mm might collect a non-representative high amount of larvae and  
 179 juveniles.

180 All gastropod and bivalve shells and shell fragments were analyzed and identified  
 181 to the species level with some exceptions of shells that were unidentifiable due to  
 182 preservation issues. Other skeletal elements not included in the study were: crab  
 183 fragments, serpulid tubes, abundant cirriped plates, sea-urchin spines, fragments

**Fig. 3.3** Sampling method:  
sieving samples



184 of bryozoan colonies, oyster recruits on large valves and polyplacophoran plates.  
 185 Cirripeds and cirriped plates, although very abundant, were excluded from analysis  
 186 due to the difficulty in identifying the plates. Gastropod shells and articulated bivalves  
 187 were assigned one count. Left and right valves of bivalve species were counted  
 188 separately. The count per species resulted from the sum of articulated specimens  
 189 plus the most abundant valves (left or right). Some oyster and *Pododesmus* shells  
 190 that could not be identified were counted together and divided by 2. Bivalve fragments  
 191 were counted if the umbo and at least one-third of the valve were preserved (very  
 192 small fragments were discarded). Gastropod fragments were counted when they  
 193 contained the apex and at least half of the shell.

### 194 3.4.3 Statistical Methods

195 Counts were made per level (LI, MI, UI, and Supra), and the coarse mesh size  
 196 fraction of samples was also registered separately for each level. Diversity was esti-  
 197 mated using different indices: species richness (S, the raw number of species and  
 198 by rarefaction), the Shannon-Wiener ( $H'$ ) index, the equitability J index (Hammer  
 199 and Harper 2006) and the probability of an interspecific encounter (PIE), an even-  
 200 ness index (Hurlbert 1971). Rarefaction to the lowest sample size was calculated in  
 201 order to evaluate species richness without the effect of sample size. The  $H'$  index  
 202 summarises information on species richness and evenness and correlates with S and  
 203 sample size, as does the J index. The PIE index was added to obtain an estimation  
 204 of evenness which was not affected by sample size (Olszewski and Kidwell 2007).

205 Data management and calculation of the PIE index according to Hurlbert's formula  
206 was carried out using standard spreadsheet software. Other diversity indices were  
207 calculated using PAST v 3.15 (Hammer et al. 2001).

208 Samples (levels) were plotted using a non-metric multidimensional scaling  
209 (NMDS) ordination analysis to evaluate their similarity. The database was first trans-  
210 formed to percentages per sample, then square root transformed, and then a similarity  
211 matrix was calculated based on the Bray-Curtis index (Clarke 1993; Clarke and War-  
212 wick 2001; Clarke et al. 2006). NMDS was carried out using R software, version  
213 3.4.3 (R Core Team 2017).

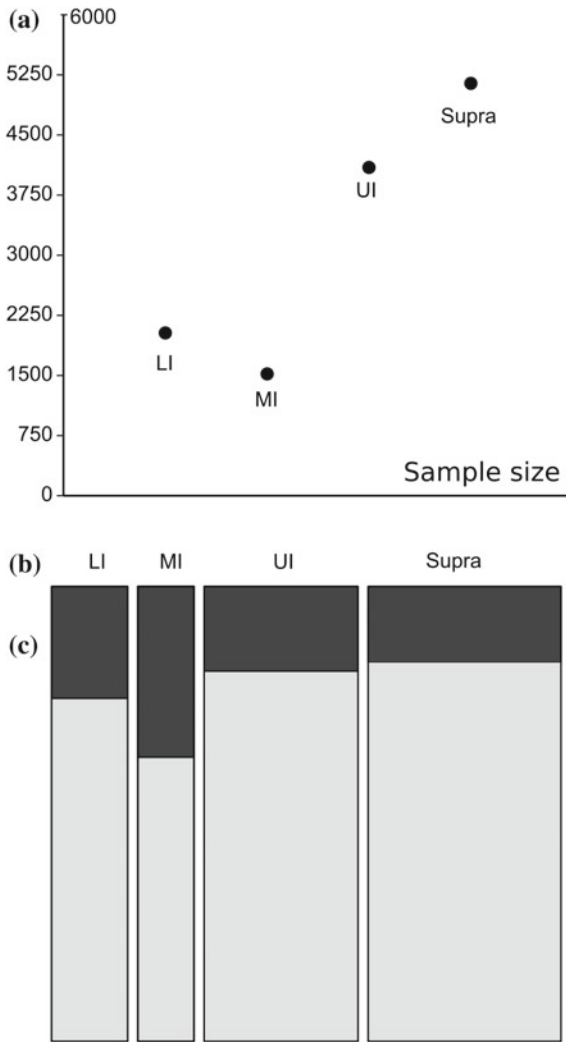
214 To test the effect of using samples sieved with coarse mesh (as proxies for col-  
215 lecting by hand), we compared these with the results obtained for whole samples  
216 (coarse + fine mesh) by using diversity indices and an ordination plot (NMDS).

217 Beta diversity was quantified in order to assess both the existence of a gradient  
218 along the coastal profile for the four levels (directional turnover) and non-directional  
219 variation for comparing the coarse mesh subsample with the complete sample (whole  
220 = coarse plus fine mesh subsamples) (Anderson et al. 2011). To evaluate the gradient  
221 in beta diversity, the similarity between the supratidal sample and the samples from  
222 all other levels was calculated with the Jaccard similarity index on a presence/absence  
223 matrix. The results were plotted in their position on the coastal profile, from Supra  
224 to LI. If species turnover along the gradient existed, then a pattern of similarity  
225 decrease would be expected from left to right. To determine the differences in species  
226 presence/absence in coarse and fine samples, Whittaker's beta diversity index ( $\beta_w$ )  
227 was calculated between pairs of coarse and whole samples per level, and then plotted  
228 in their position on the coastal profile. Higher levels of  $\beta_w$  imply a greater mismatch  
229 between the coarse mesh samples and the whole samples (Koleff et al. 2003).

### 230 3.5 Results

231 A total of 12,790 mollusk specimens belonging to 31 bivalve and 39 gastropod  
232 species were collected (Table 3.1, Supp Tables A and B). The sample size was  
233 uneven between levels due to the variable densities of shells in sediments from the  
234 different samples (Fig. 3.4a). Lower and middle intertidal samples contained less  
235 than half the specimens of the upper and supratidal samples. The coarse fraction per  
236 level fluctuated between 17 and 38% (Supra and MI respectively, Figure 3.4b). MI,  
237 with the smallest sample size (1520 specimens), has the highest percentage of the  
238 coarse fraction (38%).





**Fig. 3.4** **a** Size of sample per level. Levels, LI: lower intertidal, MI: middle intertidal, UI: upper intertidal, Supra: supratidal. n: number of specimens. **b** Size of samples per level and proportion of specimens captured in the coarse mesh. **c** coarse mesh. Width of bars express sample-size

### 3.5.1 Alpha and Beta Diversities Across the Intertidal Gradient

The 70 species identified in this study are distributed differently across samples (Tables 3.1 and 3.2). The *S* index is highest in Supra, followed by LI, UI, and MI. However, when standardizing to  $n = 1520$  by rarefaction, the highest diversity is found in LI (46), followed by Supra, MI and UI, which have between 36 and 39 species (Fig. 3.5a, b). The rarefaction curves show that none of the samples have a stabilizing size pattern (Fig. 3.5c), suggesting that larger sample sizes are necessary to accurately document the kind of study.

Evenness differs between levels, and is consistently lowest in Supra, followed by UI, and then LI and MI with higher values (Table 3.2 and Fig. 3.6a, b and c). The  $H'$  index is highest for LI, while the *J* index has MI as the evenest sample. The *PIE* index, which is more reliable for studies with different sample sizes, is highest for LI, followed by MI, UI, and Supra, coinciding with the  $H'$  index.

Multivariate ordination using an NMDS plot indicates a similarity between LI and MI, while UI and Supra remain separate (Fig. 3.7a). The analysis of beta diversity

**Table 3.1** Distribution of counts per level and mesh size

Sample type	LI		MI		UI		Supra		Total	
	C	F	C	F	C	F	C	F	C	F
<i>n</i>	2031		1520		4095		5144		12,790	
	510	1521	581	939	773	3322	885	4259	2750	10,040
<i>S</i>	49		37		45		55		70	
	37	32	29	31	36	33	34	46	53	52

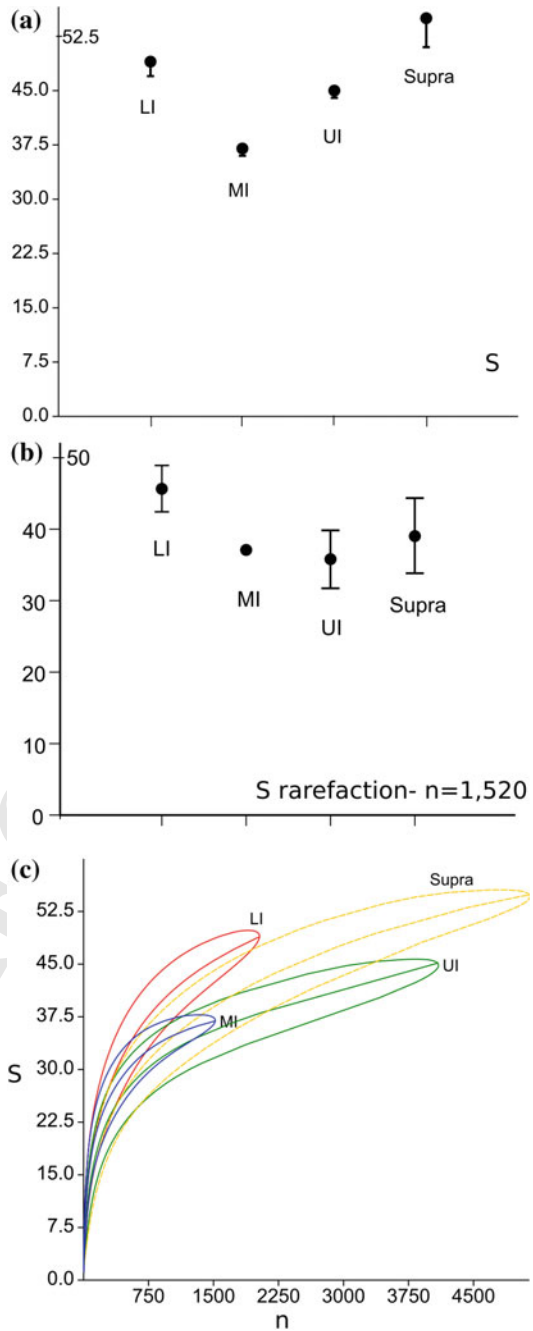
*LI* lower intertidal level, *MI* middle intertidal level, *UI* upper intertidal level, *Supra* supratidal level or high-water mark. *n* number of specimens, *s* number of species (species richness). Sample type: *C* coarse mesh, *F* fine mesh

**Table 3.2** Diversity indices calculated per level and for the pooled sample

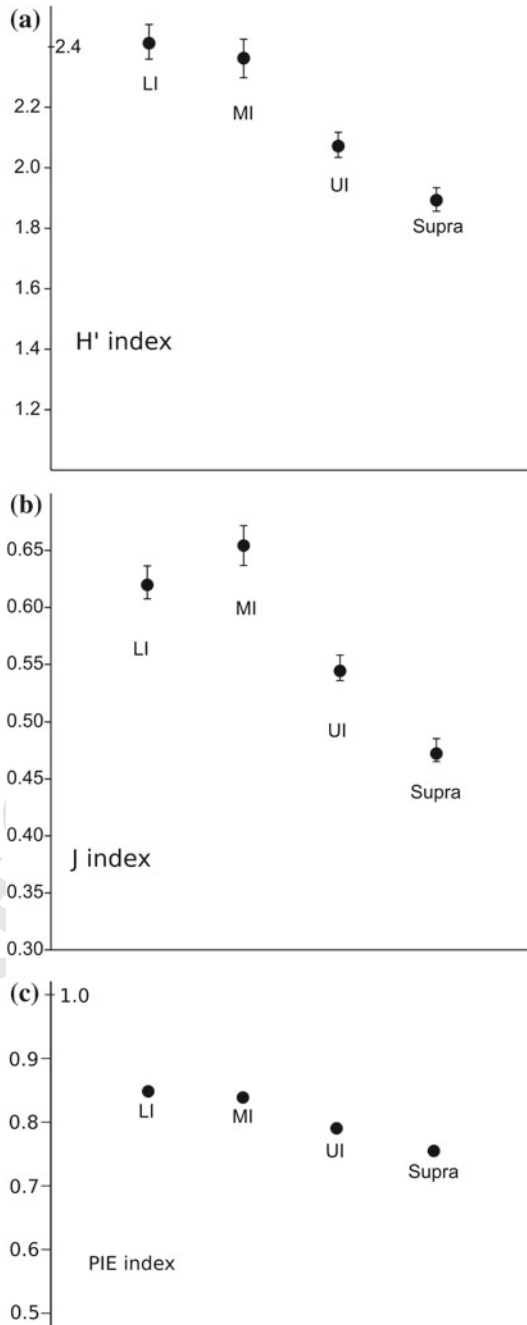
	LI	MI	UI	Supra	Pooled sample
<i>n</i>	2031	1520	4095	5144	12,790
<i>S</i>	49	37	45	55	70
<i>S</i> -rarefaction	46	37	36	39	
$H'$	2.412	2.362	2.072	1.893	2.168
<i>J</i>	0.620	0.654	0.544	0.472	0.512
<i>PIE</i>	0.847	0.838	0.791	0.755	0.799

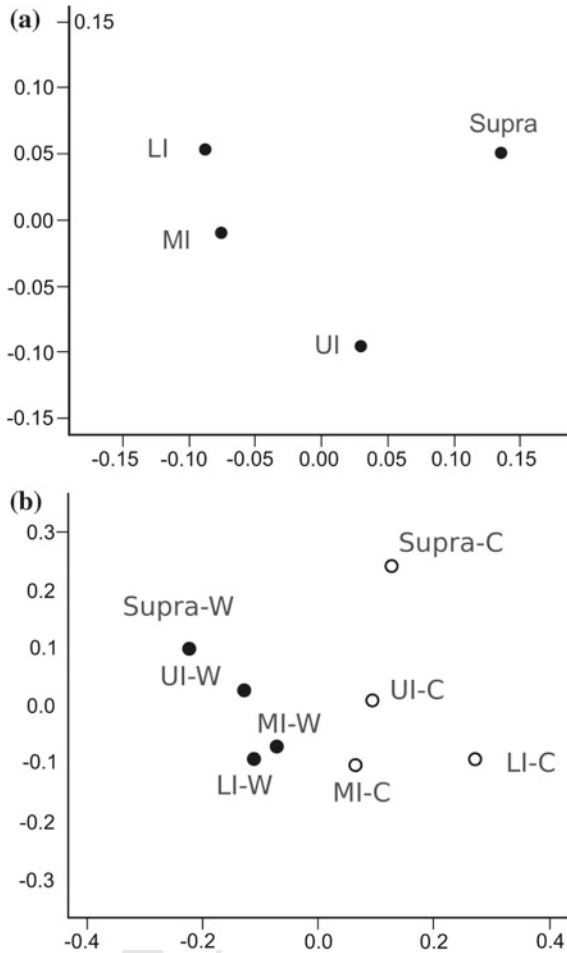
*n* sample size, *S* species richness, *S*-rarefaction species richness at a sample size of 1520,  $H'$  Shannon-Wiener index, *J* Pielou's evenness index, *PIE* probability of interspecific encounter diversity index

**Fig. 3.5** **a** Plot of species richness (S) per level. The bar represents a bootstrap 95% confidence interval. **b** Rarefaction species richness to n = 1520 per sample. The bar includes 2 standard errors. **c** Rarefaction curves per level with 95% bootstrap confidence interval. Species richness on the y axis; sample size on the x axis



**Fig. 3.6** **a** Plot of  $H'$  index per level. **b** Plot of  $J$  index per level. **c** Plot of PIE index per level





**Fig. 3.7** **a** Non-metric multidimensional scaling plot between levels. **b** Non-metric multidimensional scaling plot per level and aperture mesh size. **c** coarse mesh sample; **W**: whole sample (coarse plus fine mesh sample)

255 allows the identification of a pattern of decrease along the supratidal to the lower  
 256 intertidal gradient (Fig. 3.9a).

### 257 3.5.2 Effect of Mesh Aperture Size

258 The samples sieved with coarse mesh have richness and equitability values which  
 259 are lower than estimations for whole (coarse + fine) samples (Table 3.2 and 3.3).  
 260 Coarse mesh samples consistently underestimate the species richness of the death

**Table 3.3** Diversity indices calculated per level for the coarse aperture mesh sieved samples

Index	LI-C	MI-C	UI-C	Supra-C
n	511	581	773	885
S	37	29	36	34
S-rarefaction	37	28	32	28
H'	2.396	2.279	1.959	1.442
J	0.664	0.677	0.547	0.409
PIE	0.822	0.793	0.684	0.510

*n* sample size, *S* species richness, *S-rarefaction* species richness at a sample size of 511, *H'* Shannon-Wiener index, *J* Pielou's evenness index, *PIE* probability of interspecific encounter diversity index, *C* coarse mesh

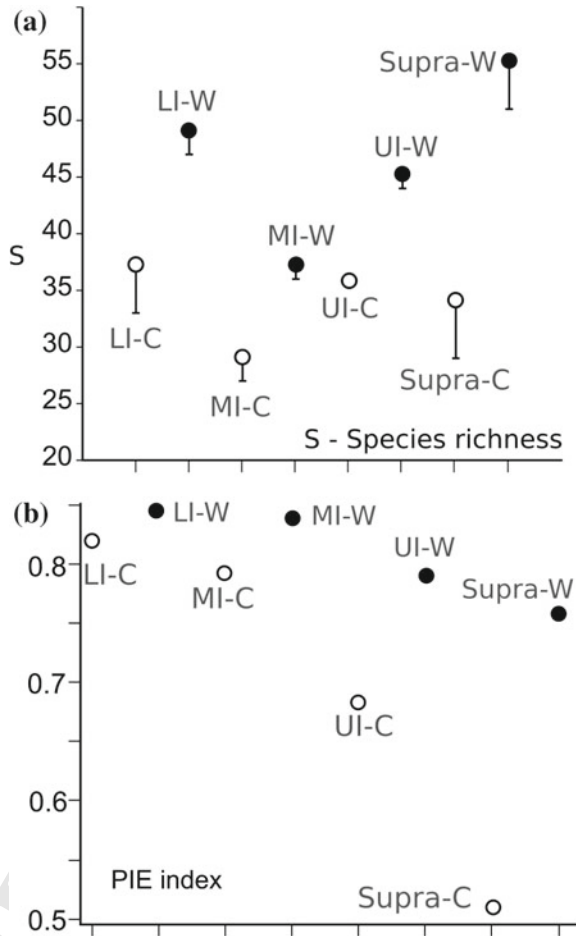
261 assemblage (Fig. 3.8a), and are less even for all four levels (PIE index, Fig. 3.8b). On  
 262 the NMDS ordination plot, coarse mesh samples cluster together and are separate  
 263 from the whole samples (Fig. 3.7b). The comparison of Whittaker's beta diversity  
 264 indices ( $\beta_w$ ) shows a large mismatch between the coarse mesh and whole samples at  
 265 the supratidal level (Fig. 3.9b), suggesting that at this level the species composition  
 266 of the coarse sample is the least similar to the whole sample.

## 267 3.6 Discussion

### 268 3.6.1 Alpha and Beta Diversity Trends

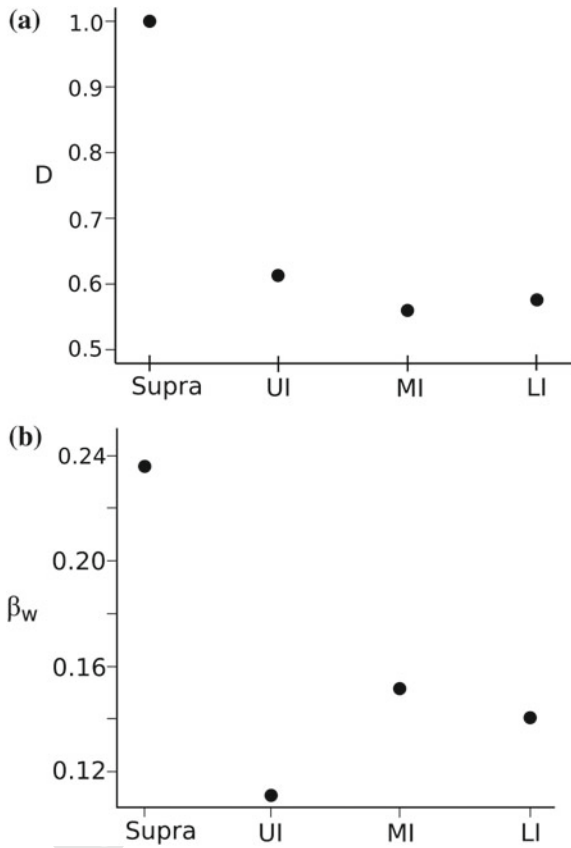
269 There is a general trend in decreasing diversity from the lower intertidal to the  
 270 supratidal belt, both for species richness and evenness. The pattern is more evident in  
 271 estimations not dependant on sample size (Figs. 3.5b and 3.6c) than in measurements  
 272 associated with sample size (Figs. 3.5a and 3.6a, b). In the case of species richness,  
 273 its estimation via rarefaction to the lowest sample size ( $n = 1520$ ) suggests a trend  
 274 from LI to UI, although the Supra sample is slightly more diverse than MI and UI  
 275 (Fig. 3.5b). The PIE index shows a decrease in evenness from LI to Supra, and a  
 276 similar situation can be observed in the *J* and *H'* indices, despite the effect of sample  
 277 size (Fig. 3.8a, b and c). NMDS ordination does not reflect a clear pattern. However,  
 278 the values of Bray-Curtis similarity indices between levels follow the LI to Supra  
 279 gradient (Table 3.4): contiguous samples are more similar to one another than non-  
 280 contiguous ones. Distance between samples was coded as 1 to 3 (1 for contiguous  
 281 samples, 2 for LI to UI and Mi to Supra; and 3 for LI to Supra), and the Spearman  
 282 rank correlation index was calculated between the Bray-Curtis index and distance,  
 283 thereby obtaining a significant negative correlation of  $-0.93$  ( $p = 0.033$ ). Additional  
 284 evidence of the influence of the depth gradient in the species composition of samples  
 285 comes from the evaluation of beta diversity: compared with the Supra level, there is

**Fig. 3.8** **a** Species richness (S) per level and discriminating coarse mesh sample (C) from whole sample (W). **b** PIE index as estimation of evenness per level and per mesh size. C: coarse mesh sample; W: whole sample (coarse plus fine mesh sample)



286 a decrease in similarity from UI to LI. This is interpreted as a consequence of the  
 287 depth gradient, whether due to a taphonomic gradient explained by biostratigraphic  
 288 factors (transport by waves and wind, selective destruction), the species composition  
 289 gradient of the living community, or both. In turn, harsh environmental conditions  
 290 (high levels of desiccation, strong winds, and wave energy) suggest that the upper  
 291 intertidal, and particularly the supratidal, belts should have poorer living community  
 292 diversity; however, this is not seen in the death assemblages, thus inferring vertical  
 293 transport from the middle and lower intertidal and shallow subtidal. The higher  
 294 than expected richness in the Supra sample might be a consequence of the trapping  
 295 (stranding) of shells above the high water mark during energetic storms; shells and  
 296 live specimens are stranded above the high water mark, and are no longer reached  
 297 by usual or normal storm waves (López et al. 2008).

**Fig. 3.9** **a** Beta diversity comparison along the supratidal to lower intertidal gradient. D: Jaccard distance index between Supra level and the other three levels. **b** Whittaker beta diversity index between whole sample and coarse mesh sub-sample per level



**Table 3.4** Bray-Curtis similarity between levels (same similarity matrix used for NMDS)

Levels compared	Type of comparison	Bray-Curtis index
LI-MI	Contiguous levels	0.869
MI-UI	Contiguous levels	0.834
UI-Supra	Contiguous levels	0.816
LI-UI	One level skipped	0.809
UI-Supra	One level skipped	0.773
LI-Supra	Two levels skipped	0.768



**Table 3.5** Bray-Curtis similarity indices of the whole sample (pooled) against samples of every level

Level	Bray-Curtis index
LI	0.858
MI	0.856
UI	0.895
Supra	0.886

298 Finally, Bray-Curtis similarity was calculated to compare each level with the  
 299 whole sample (pooled), and it was found that the Supra and UI levels are the most  
 300 similar to the total sample (Table 3.5). Although in every case abundances were  
 301 standardized to percentages and square root transformed, the most abundant samples,  
 302 Supra and UI, might still influence the result and cause this similarity. The best  
 303 sampling strategy would still be to collect material from every level, but sampling  
 304 death assemblages accumulated on the high water mark (Supra) level would not lead  
 305 to important biases. However, it must be considered that the Supra level has the  
 306 highest bias when fine meshes are not used, as seen below.

307 Whether diversity along the lower intertidal to supratidal areas of this study follows  
 308 a gradient of a biological (species composition of the living community), taphonomic  
 309 (differential transport, destruction and shell production among species) or mixed  
 310 nature, will hopefully be answered in an ongoing study which is focusing on a  
 311 level-by-level live-dead comparison. As for soft-bottom studies, there is still a need  
 312 for more actualistic updated research (Olszewski and Kidwell 2007; Kidwell 2015;  
 313 Tyler and Kowalewski 2017). Our investigation is particularly relevant because it  
 314 helps to fill the need for studies of this kind on hard-bottom environments (Smith  
 315 2008; Archuby et al. 2015).

### 316 3.6.2 Mesh Size Matters: The Effect of Sieving 317 on Biodiversity Assessment

318 There are relevant differences between the coarse fraction and the whole sample  
 319 (whole samples are composed of coarse and fine mesh samples. For an explanation,  
 320 see Sampling in Methodology). Coarse samples are less even (Fig. 3.8b) and have  
 321 lower species richness (as expected, since “coarse” samples are part of the “whole”  
 322 samples of each level; Fig. 3.8a). Species richness in coarse samples ranges from  
 323 80 to 62% of the values obtained for the whole samples, with equivalent sampling  
 324 efforts. The differences observed are also reflected in the ordination plot, in which all  
 325 coarse samples are clustered together and separated from whole samples (NMDS,  
 326 Fig. 3.7b). Discordance in terms of beta diversity is more marked in Supra than  
 327 in the rest of the samples. This means that in Supra, the mismatch between coarse  
 328 and whole samples is highest. This is still more relevant if we consider that many  
 329 minute species, such as *Phlyctiderma semiaspera*, *Turbonilla macaensis* or *Anachis*

330 *isabellei*, are present in the coarse samples because they were stuck to mytilid byssus,  
 331 which would not otherwise have been sampled by hand. Without these queue-jumper  
 332 species, the biases of coarse samples would have been even larger. Olszewski and  
 333 Kidwell (2007) detected that the evenness of death assemblages is more similar to live  
 334 assemblages when samples are sieved with a mesh size finer than 2 mm, which was  
 335 also the case for species richness. A positive bias in evenness and species richness  
 336 in coarse mesh samples with respect to fine mesh samples (which are in turn more  
 337 similar to live assemblages), can be caused by the greater durability of large mollusk  
 338 shells, lower temporal volatility of adult specimens in living communities, or both  
 339 (Kidwell 2002; Olszewski and Kidwell 2007).

340 The positive effect of sieving with fine meshes in diversity studies based on DAs is  
 341 supported by evidence from living communities. In a compilation of mollusc diversity  
 342 (including bivalves, gastropods, polyplacophorans, scaphopods and cephalopods) in  
 343 the San Matías and San José gulfs, Güller and Zelaya (2017) noted that out of the total  
 344 196 species described for the whole area, 61 (31.1%) have a maximum size smaller  
 345 than 10 mm. As a consequence, almost one-third of the species in the assemblage  
 346 have a lower probability of being collected, and would perhaps be neglected if fine  
 347 mesh size had not been used. The most abundant gastropod and bivalve species found  
 348 by these authors, respectively *Parvanachis isabellei* and *Crenella divaricata*, fit this  
 349 condition.

### 350 3.6.3 *Sampling Issues: Features of Death Assemblages Along* 351 *the Intertidal Belt*

352 Dead shell assemblages differ from living communities due to time-averaging and dif-  
 353 ferential transport and destruction (Kidwell 2001). The effect of tidal regime together  
 354 with the action of waves on the bottom differ in intensity along the intertidal belt.  
 355 As a consequence of these differences in the intensity of transport, destruction, and  
 356 sorting of shells, some variation might be expected in shell density, bioclast size, and  
 357 vertical transport, which would affect the results of sampling. Shell density varies  
 358 along the intertidal belt, as evidenced in the individuals counted per sample (Fig. 3.4a  
 359 and Table 3.1). The rarefaction curves indicated that none of the samples were large  
 360 enough to be representative of species diversity, so more sampling is therefore needed,  
 361 especially from the lower and middle intertidal levels. The proportion of coarse to  
 362 fine mesh shells varies little; the exception is MI, but the fact that this level has the  
 363 smallest sample size could explain the difference (Fig. 3.4b). Strong vertical trans-  
 364 port of shells in DAs is also evident since the species composition of all the samples  
 365 includes at least some taxa that are characteristic of subtidal to lower intertidal belts  
 366 (e.g. *Aulacomya atra*, *Venus antiqua*, some *Buccinanops* species, etc.). On the other  
 367 hand, these rocky Patagonian shores have little diversity of living fauna in the upper  
 368 intertidal belt, and almost no marine life higher up, in the supratidal fringe (Bertness  
 369 et al. 2006 and personal observations).

### 3.6.4 *Mollusc Shell Dead Shell Assemblages as Samples of Living Marine Biodiversity*

Two theoretical frameworks have come together in this study: 1. conservation biology has supplied information on the use of biological surrogates of marine benthic communities (Magierowski and Johnson 2006; Smith 2005, 2008; Mellin et al. 2011; Tyler and Kowalewski 2017); and 2. taphonomy, and the new, related discipline conservation paleobiology, have contributed with the assessment of the processes that operate between living communities and accumulations of their remains, for those taxa that bear mineralized or highly durable tissues (Kidwell 2001; Tomašových and Kidwell 2009a; Dietl and Flessa 2011). Put simply: if a particular taxon is an appropriate surrogate for a living community, and the accumulation of its durable remains or death assemblages are good proxies of the living counterpart of the taxon, then the death assemblages are highly valuable tools as rapid and faithful proxies of the living communities.

Different studies coincide on the point that exhaustively sampling living marine diversity is almost impossible, very expensive and particularly time-consuming, mainly due to the difficulties involved in accessing study sites, poor taxonomic knowledge and the high diversity of marine communities (Warwick and Light 2002; Magierowski and Johnson 2006; Smith 2008; Mellin et al. 2011; Tyler and Kowalewski 2017). Besides, results show that at least in some cases (depending on habitat type and spatial scale), mollusks are appropriate surrogates of marine communities (e.g., Smith 2008; Tyler and Kowalewski 2017). Dead shell assemblages represent time-averaged relics of the communities they come from. Their differences from living assemblages are explained mainly by their time-averaged nature: they are composed of a mixture of successive communities that lived in the same area and are modified by vertical and lateral transport and other biostratigraphic agents (Kidwell 2001, 2013; Archuby et al. 2015). One of the expectations with respect to the features of death assemblages is an increase in species richness and evenness (Tomašových and Kidwell 2010a). Olszewski and Kidwell (2007) demonstrated that on average death assemblages surpass living communities in species richness and evenness, although particular examples might have a different pattern (with little frequency). The only study of live/dead comparisons along the Patagonian Atlantic coast detected a systematic increase in both diversity measurements in every single comparison (Archuby et al. 2015), even when sediments were not sieved (just hand collected) and live and dead samples did not coincide in time or extent (living communities were only sampled from the middle intertidal a few years before the collection of death assemblages).

In order to evaluate to what degree our death assemblages can provide relevant information on regional biodiversity, we compared them with data from surveys of benthic communities. Relevant studies on living mollusc diversity in the San Matías and San José gulfs are summarised in Table 3.6, including species richness (discriminating between bivalves and gastropods), the nature of the sample (life or death assemblage), the extent of the sampling area, the sampling effort (in terms of

Table 3.6 Living and death assemblages studied in San Matías and San José Gulfs

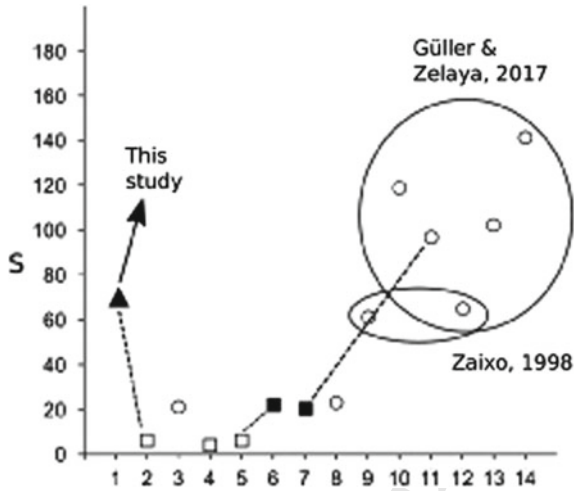
Locality	Biv	Gast	Moll	DA/LA	Extent	K	SE	n
1. PM (this study)	31	39	70	DA	Small	1	Moderate	12,681
2. PM (Archuby et al. 2015) (2)	4	2	6	LA	Small	1	Low	5573
3. SMG (Avaca et al. 2008, NOR)	12	9	21	LA	Medium	122	High	np
4. LG (Archuby et al. 2015) (2)	3	1	4	LA	Small	1	Low	3903
5. PD (Archuby et al. 2015) (2)	4	2	6	LA	Small	1	Low	3564
6. PD (Archuby et al. 2015) (1)	10	14	24	DA	Small	1	Low	686
7. PL (Archuby et al. 2015) (1)	8	13	21	DA	Small	1	Low	1826
8. SMG (Avaca et al. 2008 NOE)	19	4	23	LA	Medium	32	High	np
9. SJG, SMG. (Zaixo et al. 1998)	25	36	61	LA	Large	120	Very high	np
10. SAB (Güeller and Zelaya 2017) (3)	71	48	119	LA	Large	np	Very high	16,479 (4)
11. PL (Güeller and Zelaya 2017) (3)	54	43	97	LA	Large	np	Very high	2861 (4)
12. SJG (Güeller and Zelaya 2017) (3)	26	39	65	LA	Large	np	Very high	np
13. PD (Güeller and Zelaya 2017) (3)	58	44	102	LA	Large	np	Very high	np
14. SMG (Güeller and Zelaya 2017) (3)	60	81	141	LA	Large	85	Very high	30,481

(1) Data from Archuby et al. (2015) plus additional information that completes the death assemblage. (2) Samples taken only at the middle intertidal horizon. (3) Only bivalves and gastropods, restricted to species found alive. (4) Authors provide self sampled sizes but some species were not found but posteriorly added from literature, with no indication of additional sample sizes. *np* not provided in the source literature. Acronyms: *PM* Punta Mejillón, *SJG* San José Gulf, *SMG* San Matías Gulf, *PD* Playas Doradas, *PL* Puerto Lobos, *SAB* San Antonio Bay, *Biv*, *Gast*, *Moll*/bivalves, gastropods and molluscs respectively, *DA* death assemblage, *LA* living assemblage, *Extent* area comprised in the sampling, *K* number of samples, *SE* sampling effort, *n* number of specimens

413 cost and time of the sampling process) and, when available, sample size. Archuby  
414 et al. (2015) studied a series of live and dead mollusk assemblages, spanning 1500 km  
415 from Punta Mejillón to Puerto Deseado in the South of Patagonia. Their case studies  
416 from the San Matías Gulf were numbered 2, 4, 5 (LAs), 6 and 7 (DAs). Avaca  
417 et al. (2008) provided two databases of living communities, sampled in the North  
418 and Northwest margins of the San Matías Gulf. Case study 9 corresponds to an  
419 extensive study in the San José Gulf and the closest part of the San Matías Gulf  
420 (Zaixso et al. 1998). Recently, Güller and Zelaya (2017) published a highly-qualified  
421 study with information on mollusk species from the San Matías and San José gulfs,  
422 which includes results from their own samples and from an exhaustive bibliographic  
423 compilation. The authors used 85 sampling points from the intertidal to a depth  
424 of 25 m in the subtidal, and also took some (not detailed) samples from deeper  
425 bottoms, obtaining a total of 30,481 mollusk specimens, including empty shells and  
426 valves. Their database, and most of their study focused on four main areas which  
427 together account for 119 species of bivalves (49) and gastropods (70) for both gulfs  
428 (when considering only live species found in their samples). When other studies are  
429 added, the species count for the whole area reaches 141 species (60 bivalves and 81  
430 gastropods).

431 Death assemblages offer relatively high levels of diversity compared to life assem-  
432 blages (Fig. 3.10). Individual (single place) samples of living communities such as 2,  
433 4 and 5 represent low effort sampling but with very little diversity (up to 6 species).  
434 Several-point samples of living communities demand high levels of effort (availabil-  
435 ity of vessels, complex sampling devices, diving). Case studies 3 and 8 are samples  
436 of living communities with 122 and 32 sampling points respectively, where only  
437 21 (case study 3) and 23 (case study 8) species were collected. In these two cases,  
438 the sampling area was considered of medium size (narrow fringes parallel to the  
439 coast). Case study 9 (Zaixso et al. 1998) is a 120 sample point survey in which a  
440 species richness of 61 bivalves and gastropods was collected. Case study 12, taken  
441 from Güller and Zelaya (2017), is based on Zaxso's data plus additional information.  
442 Case studies 10, 11 and 13 are based on multi-point sampling of living communities  
443 across wide areas of the San Matías Gulf. Case study 14 represents a synthesis of  
444 multipoint sampling and all available published information on mollusk diversity  
445 in the San Matías and San José gulfs. Güller and Zelaya (2017) and Zaixso et al.  
446 (1998) sieved their samples with less than 2 mm aperture meshes, while Avaca et al.  
447 (2008) used 40 mm aperture nets. According to Güller and Zelaya (2017), sampling  
448 without fine meshes drastically reduces diversity, since they detected that more than  
449 45% of species have shells with a maximum size smaller than 15 mm. This differ-  
450 ence in sampling strategy might have caused the reduced diversity record in Avaca's  
451 samples. Güller and Zelaya (2017) consider that the lower than expected diversity  
452 found in the San José Gulf compared with their results in other areas is due to the  
453 different sampling methods used (they did not actually sample this gulf, but instead  
454 summarised information from other studies).

455 The three death assemblage case studies, 1 (this study), 6 and 7 (Archuby et al.  
456 2015), are single point samples that represent moderate or low effort but offer a  
457 relatively high number of specimens and high diversity. Case studies 6 and 7 sampled



**Fig. 3.10** Plot of calculated species richness in different case studies in the San Matías and San José gulfs. Order of case studies and acronyms of location sampling areas, as in Table 3.6. (1: PM (this study). 2: PM (Archuby et al. 2015) (2). 3: SMG (Avaca et al. 2008. NOR). 4: LG (Archuby et al. 2015) (2). 5: PD (Archuby et al. 2015) (2). 6: PD (Archuby et al. 2015). (1). 7: PL (Archuby et al. 2015) (1). 8: SMG (Avaca et al. 2008. NOE). 9: SJG, SMG. (Zaixo et al. 1998). 10: SAB Güller and Zelaya 2017 (3). 11: PL Güller and Zelaya 2017 (3). 12: SJG Güller and Zelaya 2017 (3). 13: PD Güller and Zelaya 2017 (3). 14: SMG Güller and Zelaya 2017 (3). Empty symbols: life assemblages. Filled symbols: death assemblages. Squares: low sampling effort. Triangles: moderate sampling effort. Circles: high sampling effort. Slashed lines join samples from comparable localities

24 and 21 species respectively, which is similar to the 21 and 23 species in Avaca et al. (2008) which required 122 and 32 samples for a similar result. However, those samples were taken without sieving. In our study, a single point in Punta Mejillón, sampled using  $1 \times 1$  mm aperture mesh, detected higher species richness than 122 samples throughout the entire San José Gulf.

This study has confirmed that sampling species richness requires sieving with fine mesh sieves for both living communities (Güller and Zelaya 2017) and their associated death assemblages (Kidwell 2002). In turn, death assemblages give excellent results at equivalent levels of sampling effort, if compared with the sampling of life assemblages. This can be explained by the time-averaging and spatial homogenization of successive communities which accumulated to constitute the death assemblage (Tomašových and Kidwell 2010a).

A word of caution is needed here: the case studies of living communities used for comparisons are mainly based on samples taken from subtidal soft bottoms, while our study was carried out in a rocky-bottom intertidal belt. More quantitative research is needed in order to compare life and death assemblages from equivalent habitats and to evaluate the partitioning of biodiversity along depth gradients and in different types of seafloor in the San Matías and San José gulfs.

### 3.7 Conclusions

One of the outstanding conclusions of this work is that death assemblages accumulated in rocky-bottom coastal environments are highly informative of the regional biodiversity. In this study, we showed that a single point sample contains more species than almost every study of living communities based on dozens of samples for the same region.

The lower intertidal to supratidal (high-water mark) depth gradient contains dead shell accumulations that reflect a gradient in diversity (a decrease in species richness and evenness), as well as a pattern of species turnover. The gradient in the death assemblages is mainly explained by differential transport upwards, and also by a gradient in species turnover in the living community.

Representative samples of death assemblages must be obtained with the use of fine (up to 2 mm aperture) meshes. Supratidal death assemblages, i.e., shells accumulated in the high water mark, are an acceptable proxy of the whole intertidal to subtidal assemblage if a horizon is to be chosen for sampling. The supratidal is more diverse than expected in terms of species richness, probably due to the supply of shells by strong storms, which were then trapped, out of the reach of normal waves or normal storm waves. In turn, supratidal samples show the highest bias in coarse-mesh sieved samples (equivalent to hand-collecting). Results from this study suggest that none of the levels is fully representative, and a sample which pools the different levels is recommended. A live/dead comparison is needed to assess this question in more depth. Exploring the live/dead mismatch in rocky-bottom intertidal environments is useful for improving our knowledge of benthic marine life, and for filling a void in the studies of rocky bottoms, as well as for evaluating the novel idea of conservation paleobiology with respect to the detection of diversity-altered, human-impacted ecosystems.

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